Amendments to the Specification:

The paragraphs numbering refers to the published Application 2007/0183970. This numbering differs from the numbering used in the Action.

Please replace the Paragraph [0033] with the following paragraph.

[0033] Considered herein is a method of treating a malignancy or other disease involving accumulation of activated or neoplastic granulocytes in a subject, comprising the step of administering to said subject a therapeutically effective amount of a naked and/or conjugated anti-granulocyte antibody, fusion protein, or fragment thereof of the present invention, formulated in a pharmaceutically acceptable vehicle, either alone or in combination with other therapeutic and/or diagnostic agents. Preferably, the-method a method of treating a malignancy in a subject, comprising the step of administering to said subject a therapeutically effective amount of an immunoconjugate or fragment thereof the present invention, formulated in a pharmaceutically acceptable vehicle.

Please replace the Paragraph [0091] with the following paragraph.

[0091] Host cells containing the DNA sequences encoding the MN3 MAbs or fragments thereof or antibody fusion proteins or fragments thereof of the present invention or host cells containing the vectors that contain these DNA sequences are encompassed by the present invention. Particularly useful host cells are mammalian cells, and more specifically, myeloma cell lines, such as Sp2/0, YB2/0, NS0, and CHO, such as DG-44, as discussed in more detail below. Also useful for producing monoclonal antibodies and other fusion proteins is the PER.C6® human cell line.

Please replace the Paragraph [0120] with the following paragraph.

[0120] A recombinant host may be any prokaryotic or eukaryotic cell that contains either a cloning vector or expression vector. This term also includes those prokaryotic or eukaryotic cells, as well as an transgenic animal, that have been genetically engineered to contain the cloned gene(s) in the chromosome or genome of the host cell or cells-of-the-host-cells. Suitable mammalian host cells include myeloma cells, such as SP2/0 cells, and NS0 cells, as well as Chinese Hamster Ovary (CHO) cells, hybridoma cell lines and other mammalian host cells

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useful for expressing antibodies. Also particularly useful to express MAbs and other fusion proteins, is a human cell line, PER.C6®, disclosed in WO 0063403 A2, which produces 2 to 200-fold more recombinant protein as compared to conventional mammalian cell lines, such as CHO, COS, Vero, Hela, BHK and SP2-cell lines. Special transgenic animals with a modified immune system are particularly useful for making fully human antibodies.

Please replace the Paragraph [0141] with the following paragraph.

[0141] Cell lines and culture media used in the present invention include MN3 producing hybridoma cells and Sp2/0-Ag14 myeloma cells (ATCC, Roekwille, Md: Manassas, VA). These cells may be cultured in Hybridoma serum-free media (HSFM) (life Technologies, Grand Island, N.Y.) supplemented with 10% fetal bovine serum (FBS) (Hyclone Laboratories, Logan, Utah) and antibiotics (complete media). Alternatively, they may be cultured in Dulbecco's modified Eagle's Medium (DMEM) supplemented with 10% FCS (Gibco/BRL, Gaithersburg, Mass.-MD) containing 10% of FCS and 75 µg/ml gantamicin (complete HSFM) or, where indicated, in HSFM containing only antibiotics. Selection of the transfectomas may be carried out in complete HSFM containing and appropriate cytocidal drug, such as hygromycin (hyg) and methotrexate (MTX). All cell lines are preferably maintained at 37° C. in 5% CO₂.

Please replace the Paragraph [0146] with the following paragraph.

[0146] RNA isolation, cDNA synthesis, and amplification can be carried out as follows. Total cell RNA can be prepared from a MN3 hybridoma cell line, using a total of about 10⁷ cells, according to Sambrook *et al.*, (Molecular Cloning: A Laboratory Manual, Second ed., Cold Spring Harbor Press, 1989), which is incorporated by reference. First strand cDNA can be reverse transcribed from total RNA conventionally, such as by using the SuperScript™ preamplification system (Gibco/BRL, Gaithersburg, MDd+). Briefly, in a reaction volume of 20 µl, 50 ng of random hexamer primers can be annealed to 5 µg of RNAs in the presence of 2 µl of 10X synthesis buffer [200 mM Tris-HCl (pH 8.4), 500 mM KCl, 25 mM MgCl₂, 1 mg/ml BSA], 1 µl of 10 mM dNTP mix, 2 µl of 0.1 M DTT, and 200 units of SuperScript™ reverse transcriptase. The elongation step is initially allowed to proceed at room temperature for 10 min followed by incubation at 42° C. for 50 min. The reaction can be terminated by heating the reaction mixture at 90° C, for 5 min.

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Please replace the Paragraph [0156] with the following paragraph.

[0156] Antibodies can be isolated from cell culture media as follows. Transfectoma cultures are adapted to serum-free medium. For production of chimeric antibody, cells are grown as a 500 ml culture in roller bottles using HSFM. Cultures are centrifuged and the supernatant filtered through a 0.2 micron membrane. The filtered medium is passed through a protein A column (1 x 3 cm) at a flow rate of 1 ml/min. The resin is then washed with about 10 column volumes of PBS and protein A-bound antibody is eluted from the column with 0.1 M glycine buffer (pH 3.5) containing 10 mM EDTA. Fractions of 1.0 ml are collected in tubes containing 10 µl of 3 M Tris (pH 8.6), and protein concentrations determined from the absorbencies at 280/260 nm. Peak fractions are pooled, dialyzed against PBS, and the antibody concentrated, for example, with the Centricon 30 (Amicon, Beverly, Mass.). The antibody concentration is determined by ELISA, as before, and its concentration adjusted to about 1 mg/ml using PBS. Sodium azide, 0.01% (w/v), is conveniently added to the sample as preservative.